420 Letters

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The Metabolite N-4Methoxyphenylretinamide is a Major Determinant of Fenretinide Induced Decline of Plasma Insulin-like Growth Factor-1

R. Torrisi, F. Pensa, V. Fontana, A. Costa and A. Decensi

Insulin-like growth factor (IGF-1), a potent mitogen for breast cancer cells, has been implicated in the promotional steps of breast carcinogenesis [1]. We have recently shown that the administration of the synthetic retinoid, fenretinide (N-(4-hydroxyphenyl)retinamide, 4-HPR), induces a significant decline in circulating levels of IGF-1 in breast cancer patients, which was more pronounced in younger women [2]. It is possible that the decrease of available IGF-1 in plasma might represent at least one of the mechanisms through which the retinoid interferes with the carcinogenic process. This study augments our previous work in that it endeavours to investigate the effects of plasma concentrations of 4-HPR and of its major metabolite N-(4-methoxyphenyl)retinamide (4-MPR) on plasma IGF-1.

The 20 stage I ($T_{1-2}N_0$) breast cancer patients enrolled in the study were part of a large randomised trial of contralateral breast cancer chemoprevention with 4-HPR [3]. All patients had been randomised to receive 4-HPR (R.W. Johnson Pharmaceutical Institute, Springhouse, Pennsylvania, U.S.A.) 200 mg daily, orally for 5 years with a monthly 3-day drug holiday. Median age was 53.5 years (range 39–67) and mean body mass index (BMI), expressed as weight (kg) divided by squared height (m²), was 24.32 \pm 0.6. Sixteen patients were postmenopausal and 4 premenopausal.

IGF-1 levels were measured by radioimmunoassay on plasma obtained at randomisation and after a mean interval of 11 ± 0.6 months, as previously described [2]. At baseline, plasma IGF-1 levels (mean \pm S.E.) were 154.8 \pm 11.4 ng/ml and decreased to 132.8 \pm 8.7 ng/ml after treatment (P=0.006, Student t-test). Plasma levels of 4-HPR and 4-MPR were determined by high-performance liquid chromatography (HPLC), on blood samples obtained after a median time on treatment of 32 months (range 6-45), as previously described [4]. Since plasma levels of the drug and the metabolite have been shown to be stable during

Correspondence to A. Decensi, Servizio di Oncologia Medica II, Istituto Nazionale per la Ricerca sul Cancro, v.le Benedetto XV 10, 16132 Genova, Italy.

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treatment for as long as 5 years [4], we assumed that the levels of 4-HPR and 4-MPR at the time of IGF-1 determination, i.e. after 1 year of treatment were comparable with those obtained at the time of our assay. Plasma levels of 4-HPR were 387.2 \pm 26.9 ng/ml and those of 4-MPR 285.2 \pm 21.7 ng/ml. When the difference (\triangle) in IGF-1 levels between follow-up and baseline values was employed as a dependent variable, simple linear regression analysis showed a significant inverse relationship of \triangle IGF-1 with 4-MPR, but not with 4-HPR levels (r=0.47, P<0.05 and r=0.08, P= not significant, respectively).

Multiple regression analysis using \(\triangle \) IGF-1 as a dependent variable and age, menstrual status, BMI, 4-HPR and 4-MPR concentrations as covariates showed a significant regression of \(\triangle IGF-1 \) on metabolite and, to a lesser extent, on drug concentrations (F test=5.50, r^2 =0.66, P<0.01). The dependence of \(\triangle IGF-1 \) on 4-MPR and 4-HPR levels was further explained by a significant interaction with age (see legend to Figure 1). In fact, when patients were categorised according to median 4-MPR concentration, in younger women the decline of IGF-1 (i.e., negative \triangle) was greater in patients with low 4-MPR levels, while in older women a negative \triangle was observed in patients with higher 4-MPR concentrations (Figure 1b). An opposite trend was observed when patients were categorised according to median 4-HPR concentration (Figure 1a). Interestingly, the occurrence of a negative \triangle appears to be constant with age in patients with higher levels of 4-MPR while a trend towards a positive \(\triangle IGF-1 \) with increasing age is observed at lower 4-MPR concentrations. Menstrual status and BMI did not affect \triangle IGF-1 (data not shown).

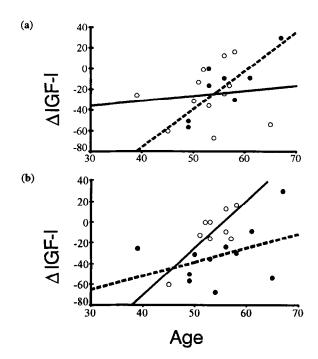


Figure 1. (a) Age-dependent behaviour of the difference (△) of plasma IGF-1 (ng/ml) in patients with plasma 4-HPR above (♠,---) or below (○,---) the median value =370 ng/ml, β=0.01, 95% confidence interval =0.0008-0.02, P=0.04; (b) age-dependent behaviour of the △ IGF-1 in patients with plasma 4-MPR above (♠,---) or below (○,---) the median value =290 ng/ml, β= -0.04, 95% confidence interval=0.07-0.01, P=0.01 (see text for detailed explanation). Data are expressed as expected values.

The authors are at the Department of Medical Oncology II (RT, FP, AD) and of Biostatistics (VF), National Cancer Institute for Cancer Research, Genoa and Division of Senology, European Institute of Oncology (AC), Milan, Italy.

Letters 421

Our results seem to suggest that 4-MPR is the major determinant of plasma IGF-1 decline (negative \triangle). This is in keeping with our previous observations of a dominant role of 4-MPR in determining another principal effect of fenretinide administration, namely, the decline of plasma retinol levels [5]. We may also speculate that the reversal of effect induced by 4-HPR and 4-MPR concentrations on \triangle IGF-1 as a function of age, is due to age-related differences in the metabolism and in tissue distribution of the two compounds, which are partially different both in mice [6] and in human mammary gland [7]. 4-MPR appears to be less extensively metabolised than 4-HPR and selectively concentrated in adipose tissue from which it may be slowly released [7]. In humans, 4-MPR has a longer half-life than 4-HPR [4], potentially exerting a prolonged effect in circulation, while having the same potency as 4-HPR in in vitro differentiation assays [8]. In addition, the metabolism to 4-MPR has recently been shown to be critical to the antiproliferative effect of 4-HPR on the growth of breast cancer cell lines [9]. Thus, the preferential effect of 4-MPR on IGF-1 may have a pharmacological explanation or, alternatively, be the result of a selective biological action elicited by 4-MPR itself, supporting a leading role for 4-MPR in determining some of the main biological effects induced in vivo by treatment with fenretinide.

1. Yee D, Paik S, Lebovic GS, et al. Analysis of insulin-like growth factor 1 gene expression in malignancy: evidence for a paracrine role in human breast cancer. Mol Endocrinol 1989, 3, 509-517.

- Torrisi R, Pensa F, Orengo MA, et al. The synthetic retinoid fenretinide lowers plasma insulin-like growth factor 1 levels in breast cancer patients. Cancer Res 1993, 53, 4769-4771.
- Veronesi U, De Palo G, Costa A, et al. Chemoprevention of breast cancer with retinoids. Natl Cancer Inst Monograph 1992, 12, 93-97.
- Formelli F, Clerici M, Campa T, et al. Five-year administration of fenretinide: pharmacokinetics and effects on plasma retinol concentrations. J Clin Oncol 1993, 11, 2036–2042.
- Torrisi R, Parodi S, Fontana V, et al. Factors affecting plasma retinol decline during long term administration of the synthetic retinoid fenretinide in breast cancer patients. Cancer Epidemiol Biomarkers Prev 1994, 3, 507-510.
- Hultin TA, Filla MS, McCormick DL. Distribution and metabolism of the retinoid N-(4-methoxyphenyl)-all-trans-retinamide, the major metabolite of N-(4-hydroxyphenyl)-all-trans-retinamide, in female mice. Drug Metab Dispos 1990, 18, 175-179.
- Mehta RG, Moon RC, Hawthorne M, et al. Distribution of fenretinide in the mammary gland of breast cancer patients. Eur J Cancer 1991, 27, 138-141.
- Swanson BN, Newton DL, Roller PP, Sporn MB. Biotransformation and biological activity of N-(4-hydroxyphenyl)retinamide derivatives in rodents. J Pharmacol Exp Ther 1981, 219, 632-637.
- Mehta RR, Mehta RG, Hart GD, Hawthorne M, Moon RC, Das Gupta TK. Effect of 4-HPR on human breast carcinoma cells. Breast Cancer Res Treat 1993, 27, A144.

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Fotemustine and Tamoxifen Combination Therapy in Metastatic Malignant Melanoma. A Phase II Study

C.J.A. Punt, J.H. Tytgat, P.A. van Liessum and B. Gerard

SEVERAL STUDIES have shown that the addition of tamoxifen to chemotherapy may enhance the response rate in patients with metastatic malignant melanoma [1-4]. Fotemustine, a nitrosurea, has activity against melanoma as a single agent [5-7], with a response rate of 24.2% in the largest study with 153 patients [5]. Tamoxifen enhances in vitro the cytotoxic effect of fotemustine on melanoma cell lines expressing oestrogen receptors [8]. In patients with metastatic melanoma, high-dose tamoxifen may result in a higher complete response rate compared with low-dose tamoxifen [9]. We, therefore, initiated a phase II study of high-dose tamoxifen and fotemustine in patients with metastatic melanoma.

Eligibility criteria included histologically confirmed metastatic melanoma, measurable progressive disease, age 18-75 years, WHO performance status ≤ 2 , life expectancy ≥ 3

Table 1. Patients' characteristics

	No. of patients
Male/female	8/5
Median age (range)	50 years (33-72)
Median WHO performance (range)	1 (0-1)
Previous therapy	
Radiotherapy	3
Chemotherapy	4
Immunotherapy	2
Number of metastatic sites	
1	5
2	1
≥ 3	7
Metastatic sites	
Cerebral	3
Visceral	10
Non-visceral	10

Correspondence to C.J.A. Punt at the University of Nijmegen, Department of Medical Oncology, P.O. Box 9101, 6500 HB Nijmegen, The Netherlands.

J.H. Tytgat is at the H. Hart Hospital, Department of Internal Medicine, Roeselare, Belgium; P.A. van Liessum is at the St. Carolus Hospital, Department of Internal Medicine, 's Hertogenbosch, The Netherlands; and B. Gerard is at I.R.I.S., Brussels, Belgium.

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